

Targeting the TGF β signaling network in human neoplasia

Nancy Dumont¹ and Carlos L. Arteaga^{1,2,3,*}

¹Department of Cancer Biology

²Department of Medicine

Vanderbilt University School of Medicine

³Breast Cancer Research Program, Vanderbilt-Ingram Comprehensive Cancer Center

Vanderbilt University Medical Center, 2220 Pierce Avenue, 777 PRB, Nashville, Tennessee 37232

*Correspondence: carlos.arteaga@vanderbilt.edu

Introduction

The transforming growth factor (TGF) β s are polypeptides within a superfamily of ligands that includes the TGF β s, activins, and bone morphogenetic proteins (BMPs). TGF β ligands play a role in cell proliferation, functional differentiation, extracellular matrix (ECM) production, cell motility, and apoptosis (Massagué, 1998). Three mammalian TGF β isoforms, TGF β 1, TGF β 2, and TGF β 3, have been identified. In general, they exhibit similar functions in vitro, most notably on cell growth regulation, ECM production, and immune modulation. However, each appears to have distinct activities in vivo, as evidenced by the distinct phenotypes of mice lacking any one of the TGF β ligands (Massagué, 1998). The TGF β s bind to a heteromeric complex of transmembrane serine/threonine kinases, the type I and type II receptors (T β RI and T β RII). Following ligand binding to T β RII, T β RI is recruited to the complex. This allows for the constitutively active T β RII kinase to transphosphorylate and activate the T β RI kinase which, in turn, phosphorylates Smad2 and Smad3. Smad2/3 then associate with a common mediator Smad, Smad4, and translocate to the nucleus, where they regulate gene transcription. By contrast, the inhibitory Smad7 can interact with T β RI and prevent the phosphorylation of the effectors Smad2/3 (Massagué et al., 2000). In addition to Smads, more recently, other signaling pathways have been implicated in TGF β actions. These include the extracellular signal-regulated kinase (ERK), c-Jun NH₂-terminal kinase (JNK), p38 mitogen-activated protein kinase (MAPK), phosphatidylinositol-3 kinase (PI3K), and Rho GTPases (reviewed in Derynck et al., 2001).

TGF β is both a tumor suppressor and a tumor promoter

The ability of TGF β to inhibit epithelial cell proliferation suggests a role in tumor suppression. MMTV/TGF β 1^{S223/225} transgenic mice expressing active TGF β 1 in the mammary gland exhibit a hypoplastic ductal epithelium (Pierce et al., 1993). Conversely, expression of dominant negative T β RII results in accelerated lobulo-alveolar mammary development (Gorska et al., 1998) and enhanced propensity for carcinogen-induced lung, mammary, and skin tumors (Bottinger et al., 1997). Mice with targeted disruption of either the *Tgfb1* or the *Smad3* gene develop colon adenomas and carcinomas (Engle et al., 1999; Zhu et al., 1998). Mice carrying an inactivated allele of *Smad4* develop intestinal polyps that progress to carcinomas (Xu et al., 2000). Finally, mice heterozygous for the *Tgfb1* gene express 10%–30% of the TGF β 1 protein, and this partial loss results in enhanced carcinogen-induced tumors (Tang et al., 1998).

TGF β function is also compromised in some human cancers. Mutations in the *TGFBR2* gene occur in both sporadic and inherited colon cancers with microsatellite instability, and

reconstitution of T β RII expression reverses transformation in colon cancer cell lines (Markowitz et al., 1995). Inactivating mutations in genes encoding Smad2 and Smad4 have been found in cancers of the gastrointestinal tract and pancreas (Hahn et al., 1996). A low level of T β RII protein identifies a cohort of women with mammary epithelial hyperplasia at increased risk for development of breast cancer (Gobbi et al., 1999). Other studies have suggested that an excess of endogenous TGF β can prevent tumor development. For example, MMTV/TGF β 1 mice are resistant to DMBA-induced mammary tumors, and MMTV/TGF α \times MMTV/TGF β 1 bigenic mice fail to develop mammary carcinomas seen in MMTV/TGF α mice (Pierce et al., 1995). Mice overexpressing active TGF β 1 under the control of the K14 promoter are relatively protected from skin tumors induced by chemical carcinogens (Cui et al., 1996). This is consistent with the ability of TGF β to maintain tissue architecture, inhibit genomic instability, and induce replicative senescence and apoptosis in nontransformed cells and/or tissues (illustrated in Figure 1). These studies support the tumor-suppressive role of endogenous TGF β . To date, however, administration of exogenous TGF β has not been reported to prevent cancer formation.

Most cancers retain TGF β receptors but attenuate TGF β -mediated antimitogenic effects. In contrast to colorectal and pancreatic cancers, Smad2 and Smad4 mutations are rare in other tumor types. Furthermore, the tumors that develop in TGF β haploid mice do not lose the second TGF β allele and, in fact, express higher levels of TGF β protein than TGF β ^{+/+} tumors (Tang et al., 1998). In addition, many tumor cells exhibit increased invasiveness in response to TGF β (reviewed in Dumont and Arteaga, 2000). TGF β can also induce an epithelial-to-mesenchymal transition (EMT) in tumor and nontumorigenic immortalized cells (Miettinen et al., 1994). Reexpression of T β RII in colon cancer cells with low invasive potential restores tumor cells invasiveness (Oft et al., 1998), and induction of TGF β 1 in papillomas induces metastatic carcinomas (Weeks et al., 2001). Patients whose pancreatic cancers overexpressed T β RII mRNA had a shorter survival than patients bearing cancers with lower T β RII mRNA (Wagner et al., 1999). Forced expression of active Smad2 in squamous cancer cells results in enhanced tumor cell motility and metastatic dissemination (Oft et al., 2002). Further, introduction of dominant negative T β RII into a variety of metastatic carcinoma cells prevents conversion to a mesenchymal phenotype and inhibits motility, tumorigenicity, and metastases (reviewed in Dumont and Arteaga, 2000). These data suggest that, although partial loss of TGF β signals is permissive for tumor development, there is no selective pressure in cancer cells to lose TGF β signaling completely. Secondly, some residual TGF β might be required for

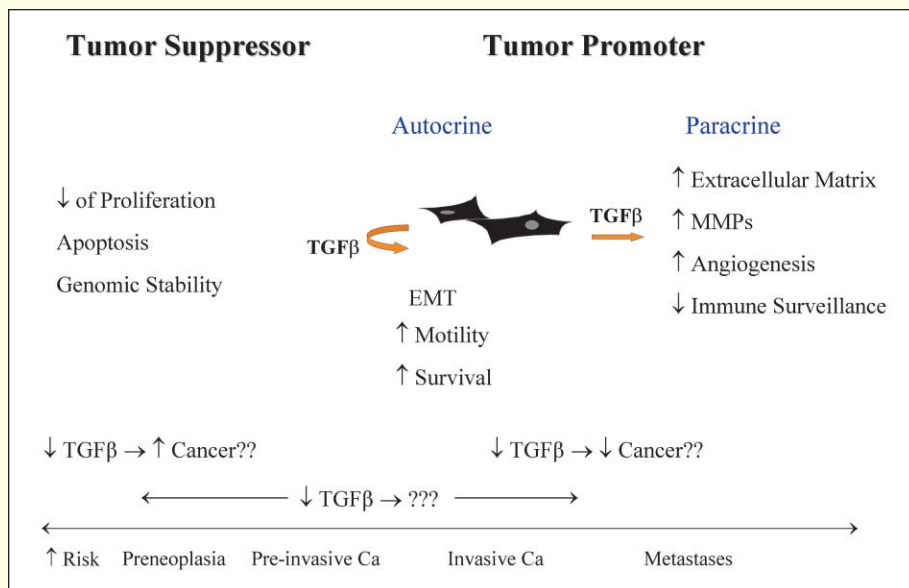


Figure 1. Diagram of tumor suppressor and tumor promoting effects of TGF β

As a result of oncogenic stimuli or other unknown mechanisms, human cancers attenuate TGF β -induced suppression but exploit TGF β -mediated autocrine and paracrine mechanisms conducive to tumor progression. It is likely that in many cancers, both situations are operative, i.e., TGF β may still attenuate proliferation while inducing cellular events associated with metastatic dissemination. There is experimental evidence that (1) loss of TGF β signaling will increase the risk of cancer development (tumor suppressor function), and (2) inhibition of autocrine/paracrine TGF β in tumors will delay metastatic progression (tumor promoter function). In the natural progression of different cancers from preneoplasia to advanced metastatic states, the timing at which the role of TGF β switches from tumor suppressor to promoter is unclear.

tumor maintenance and, in fact, may select for more metastatic cancers. Perhaps the best demonstration of this hypothesis was provided by Cui et al. (1996). Mice overexpressing active TGF β 1 in keratinocytes develop fewer benign papillomas than controls. However, the transgenic tumors rapidly acquire a spindle cell phenotype, overexpress TGF β 3, and metastasize (Cui et al., 1996). Parenthetically, colon cancers with microsatellite instability and inactivating mutations of the *TGFBR2* gene exhibit favorable survival compared to T β RII-positive colon cancers (Watanabe et al., 2001), suggesting that loss of TGF β signaling may limit systemic metastases.

There is also abundant evidence to support that excess production and/or activation of TGF β in tumors can foster cancer progression by paracrine mechanisms (Figure 1). These include an increase in tumor neoangiogenesis and extracellular matrix production, upregulation of peritumor metalloproteases, and the subversion of immune surveillance mechanisms (Derynck et al., 2001). Overexpression of TGF β ligands has been reported in most tumor types (reviewed in Wojtowicz-Praga, 2003). Most of these reports are immunohistochemical studies utilizing antibodies that do not discriminate between active and latent TGF β . Although some studies claim excess activation of TGF β in cancer compared to nontumor tissues, this can only be confirmed by use of TGF β inhibitors and assessment of their effects against TGF β -expressing tumors. Nonetheless, elevated levels of TGF β in tumor tissues correlate with markers of a more metastatic phenotype and/or poor patient outcome (reviewed in Dumont and Arteaga, 2000). In glioblastomas, TGF β 2 overexpression has been associated with immunosuppression in patients (Kuppnier et al., 1988). Elevated levels of plasma TGF β are detected in patients with cancer, and these can predict for early metastatic recurrences (Tsushima et al., 2001).

Is there a role for TGF β in cancer prevention/treatment?

A recent report indicates that a T29→C polymorphism in the *TGFBI* gene results in increased serum levels of TGF β 1 and is associated with a reduced risk of breast cancer (Ziv et al., 2001). These results plus the ability of TGF β to inhibit epithelial

proliferation and prevent tumor development in transgenic mice suggest that exogenous TGF β can be utilized for cancer prevention. However, the demonstrations that TGF β can induce signaling pathways associated with transformations like Erk, p38 MAPK, PI3K/Akt, JNK, and the Rho family of GTPases suggest that exogenous TGF β might be counterproductive as a therapeutic strategy. Furthermore, overexpression of Ras in nontumor epithelial cells synergizes with TGF β in the induction of epithelial plasticity and enhanced cell survival (Janda et al., 2002; Oft et al., 1996), suggesting that some oncogenes can potentially synergize with TGF β in early phases of transformation. Further, MMTV/neu \times TGF β 1 and MMTV/PyV mT \times TGF β 1 bigenic mice develop mammary tumors with similar latency to MMTV/neu and MMTV/PyV mT mice, respectively (C.L.A., unpublished data), implying that some oncogenes, unlike TGF α , are dominant over TGF β .

Exogenous TGF β may also have some negative consequences in patients with cancer. For example, administration of recombinant TGF β 1 to mice bearing MDA-231 xenografts does not inhibit tumor growth, but induces cachexia and generalized interstitial fibrosis in the spleen (Zugmaier et al., 1991). Daily injections of recombinant TGF β 1 facilitate tumor formation by estrogen-dependent MCF-7 human breast cancer cells in the absence of estrogen supplementation (Arteaga et al., 1993a). These data suggest the possibility of significant toxicities and a potential tumor-permissive role for treatment with this cytokine. Thus, the use of TGF β as a cancer prevention or treatment strategy is highly questionable.

Blockade of TGF β : Cellular mechanisms of action

The improved outcome of patients bearing cancers with *TGFBR2* mutations suggests an argument in favor of therapeutic blockade of autocrine TGF β signaling. An additional argument is inferred by the paracrine effects of tumor TGF β s on angiogenesis, stroma formation and remodeling, and immunosuppression. Thus, by blocking TGF β function, one can interrupt multiple events important for tumor maintenance. Indeed, preclinical studies have proved the principle that inhibition of TGF β affects these tumor-permissive autocrine and paracrine mechanisms.

Table 1. Current status of development of TGF β inhibitors

Agent	Class	Development stage	Source	References
CAT-192	Humanized TGF β 1 mAb	Phase II (diffuse systemic sclerosis)	Genzyme/CAT	
CAT-152	Humanized TGF β 2 mAb	Phase III (glaucoma)	Genzyme/CAT	Siriwardena et al., 2002
1D11	TGF β 1, 2, 3 mAb	Preclinical	Genzyme/CAT	Ananth et al., 1999
2G7	TGF β 1, 2, 3 monoclonal IgG ₂	Preclinical	Genentech	Arteaga et al., 1993b
sT β RII:Fc	RII/Fc hu IgG ₁ fusion protein	Preclinical	Biogen	Muraoka et al., 2002
Betaglycan	Recombinant soluble T β RIII	Preclinical		Bandyopadhyay et al., 1999, 2002
SB-431542	Small molecule T β RI kinase inhibitor	Preclinical	GlaxoSmithKline	Laping et al., 2002
NPC 30345	Small molecule T β RI kinase inhibitor	Preclinical	Scios, Inc.	
LY364947	Small molecule T β RI kinase inhibitor	Preclinical	Lilly Research	Sawyer et al., 2003
AP-12009	Antisense TGF β 2	Phase I/II (glioblastoma)	Antisense Pharma	Marzo et al., 1997

In MMTV/PyV mT transgenic mice, blockade of TGF β with soluble T β RII:Fc increases apoptosis in primary mammary tumors and inhibits tumor cell motility, intravasation, and metastases (Muraoka et al., 2002). In this report, treatment with sT β RII:Fc inhibited Akt activity in tumors. TGF β antibodies also inhibit PI3K/Akt activity in 4T1 and EMT-6 mammary cancer cells (Bakin et al., 2000). In transformed cells, TGF β -induced survival is associated with Akt-induced phosphorylation of FKHL1, and this effect is blocked by expression of kinase-dead Akt (Shin et al., 2001). These data imply that TGF β antagonists may need to inhibit PI3K/Akt to induce an antitumor effect. In a recent study, reconstitution of T β RI signaling in MDA-231 human breast cancer cells expressing dominant negative T β RII restored motility and enhanced survival while increasing Akt and ERK activities but not Smad2 phosphorylation (Dumont et al., 2003). This report suggests that, in some metastatic cancer cells, the tumor progressive effects of TGF β occur at a low threshold of receptor activation and may use Smad-indepen-

dent mechanisms. The clinical implications of these data are that (1) TGF β receptor-positive, Smad mutant cancers may still use TGF β for tumor progression, and (2) therapeutically, a more complete inhibition of TGF β signaling might be required to block the tumor-permissive effects of TGF β .

A monoclonal pan-TGF β neutralizing antibody suppresses the growth of TGF β 1-overexpressing renal cancer xenografts while abrogating factor VIII expression (Ananth et al., 1999), suggesting an antiangiogenic effect. Cancer cells stably transfected with soluble forms of T β RII or T β RIII exhibit reduced tumorigenicity and microvessel density compared to control tumors (Bandyopadhyay et al., 1999; Rowland-Goldsmith et al., 2001). There is also evidence that overexpression of sT β RII:Fc in thymoma cells can prevent the progression of unmodified thymoma cells when injected near the primary tumor inoculation site. Stable transduction of breast and glioma tumor cells with antisense TGF β 1 and TGF β 2 retroviruses, respectively, restores their immunogenicity when injected into immunocompetent mice which, in turn, induces partial rejection of unmodified established tumors (reviewed in Dumont and Arteaga, 2000). These examples suggest that presence of TGF β neutralizing activity within the tumor milieu can restore tumor-specific cellular immunity and mediate tumor rejection. Along those lines, transgenic mice expressing a dominant negative T β RII under the control of a T cell specific promoter eradicate tumors when challenged with live tumor cells (Gorelik and Flavell, 2001). Finally, the 2G7 pan-TGF β neutralizing IgG₂ suppresses the establishment of MDA-231 tumors and lung metastases in athymic mice and prevents the inhibition of host NK cell function induced by tumor inoculation (Arteaga et al., 1993b). The antibody had no effect against MDA-231 cells in vitro, nor did it inhibit primary and metastatic tumors in NK-deficient mice, suggesting that antibody-mediated TGF β blockade is effective in disrupting tumor-host immunosuppressive interactions that are essential for cancer progression.

Blockade of TGF β in normal cells may induce side effects in the tumor host that can be anticipated. Complete loss of TGF β in mice is associated with a severe inflammatory response. Complete elimination of TGF β function in T cells leads to autoimmune disorders (Gorelik and Flavell, 2001). A recent experiment, however, suggests that TGF β antagonists might be well tolerated. Mice expressing soluble T β RII under the regulation of the MMTV/LTR promoter exhibit high levels of the TGF β antagonist in the circulation without the severe inflammatory phenotype of TGF β null mice. The circulating levels of sT β RII:Fc were sufficient to inhibit tumor metastases in this model. Mild

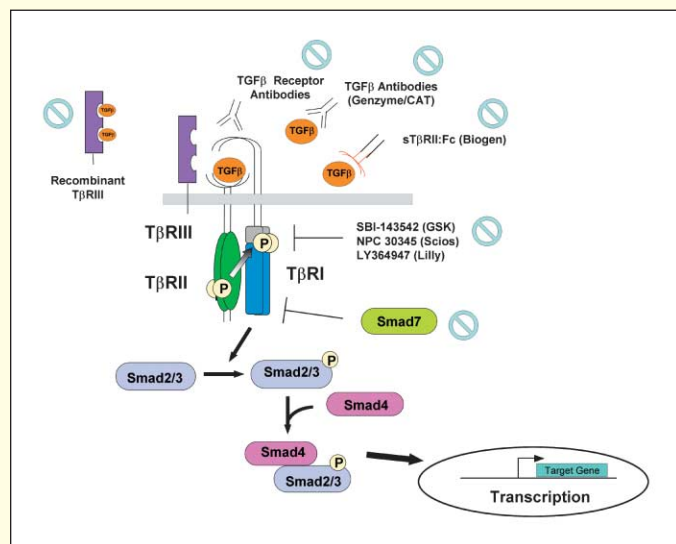


Figure 2. Sites of action of TGF β antagonists currently in development

Each class of inhibitors is marked with Ø. Smad7 is a negative regulator of TGF β signaling that has also been proposed as a therapeutic approach. There are no reports of ligand-blocking TGF β receptor antibodies in clinical development.

lymphocytic infiltration in lungs, kidneys, and pancreas were observed (Yang et al., 2002). It is likely that the severe inflammatory and autoimmune phenotype observed in genetically engineered mice reflects the complete loss of TGF β function, a level of inhibition that is unlikely to be achieved with exogenous inhibitors. Because of the stabilizing effect of TGF β on blood vessels, TGF β antagonists may affect wound healing. Interestingly, administration of a TGF β 2 antibody via subconjunctival injections immediately before and after trabeculectomy for glaucoma was not associated with impaired healing (Siriwardena et al., 2002).

Perhaps the greatest risk of TGF β antagonists is the potential acceleration of preneoplastic lesions or cancers in which TGF β still exerts growth restraint (Figure 1). In support of this possibility, expression of an antisense TGF β 1 expression vector in well-differentiated colon cancer cells enhances their tumorigenicity in nude mice (Wu et al., 1993). In other reports, administration of a TGF β neutralizing antibody or expression of soluble T β RII results in enhanced angiogenesis and/or larger primary tumor size (Gohongi et al., 1999). The possibility that blockade of TGF β with exogenous antagonists can accelerate transformation, tumor onset, and/or cancer progression requires rigorous investigation in preclinical models.

Clinical development of TGF β inhibitors

The development of anticancer molecular therapeutics requires the identification of a measurable molecular target that can aid in the selection of appropriate patients for clinical trials. This issue is not clear with TGF β antagonists, since the target network includes three ubiquitous ligands in an unknown state of activation and receptors present in both tumor and nontumor cells. The ubiquitous expression of all three TGF β ligands in human tumors, plus the inability to accurately ascertain their level of a possibly dynamic and spatially regulated activation *in situ*, argue against TGF β expression being such predictive marker for patient selection. One possible exception are patients with glioblastoma in whom TGF β 2 overexpression is associated with immune suppression. Several arguments suggest that expression of T β RII might be a good marker to use. First, most tumors retain T β RII. Second, interference of T β RII function in mesenchymally differentiated carcinomas has been shown to reverse their mesenchymal phenotype and/or inhibit metastases. Third, expression of T β RII in tumors has been associated with shorter patient survival (Wagner et al., 1999; Watanabe et al., 2001). Therefore, we propose that patients with poorly differentiated carcinomas (i.e., with mesenchymal features) that express T β RII should be appropriate candidates for the testing of TGF β antagonists. In these tumors, TGF β blockade could potentially inhibit both autocrine TGF β signaling in tumor cells and the paracrine/endocrine effects of TGF β .

Several approaches to block TGF β function are being pursued (Table 1 and Figure 2). One approach is aimed at blocking ligand access to TGF β receptors. Two humanized monoclonal antibodies: CAT-192, specific to TGF β 1, and CAT-152, against TGF β 2, are in early clinical development. The expression of multiple TGF β isoforms in tumors suggests that a pan-TGF β antibody might be more effective than isoform-specific antibodies. Two pan-TGF β monoclonal antibodies, 1D11 and 2G7, have been reported. One approach to prevent binding of TGF β ligands is the use of recombinant fusion proteins containing the

ectodomains of T β RII and T β RIII. Soluble T β RII:Fc has shown efficacy in fibrosis and metastases models. Human recombinant T β RIII has shown antimetastatic and antiendothelial cell activity. One attractive feature of recombinant betaglycan over sT β RII is its greater affinity for TGF β 2.

A second group of strategies is aimed at directly blocking the receptors' catalytic activity. SBI-14352, NPC 30345, and LY364947 are ATP competitive inhibitors of the ATP binding site of the T β RI kinase. This approach spares the T β RII kinase and, therefore, may not inhibit TGF β function completely. If complete inhibition of TGF β was required for antitumor action, this selectivity could compromise anticancer activity but at the same time ameliorate potential toxicities. These two possibilities are theoretical, since there are no known T β RII functions that do not require T β RI. Nonetheless, the development of bifunctional receptor inhibitors should help in resolving these questions. Vectors encoding the inhibitory Smad, Smad7, have been used to bind T β RI and interfere with Smad2/3 phosphorylation (Nakao et al., 1997). This strategy should be viewed with caution in that it will not block Smad-independent TGF β -induced responses conducive to tumor progression. Indeed, Smad7 mRNA is overexpressed in pancreatic cancers, and its forced expression in pancreatic cancer cells results in loss of TGF β -mediated growth inhibition but facilitates anchorage-independent growth and tumorigenicity (Kleeff et al., 1999). Moreover, blockade of Smad4 has been shown to facilitate TGF β -dependent and -independent activation of Erk in squamous cancer cells and promote their motility and transmesenchymal differentiation (Iglesias et al., 2000), suggesting that postreceptor interference of Smads as an anticancer strategy may be conceptually weak. The TGF β antagonists discussed above are summarized in Table 1.

Conclusions

There is preponderant experimental evidence to suggest that TGF β signaling can foster tumor-host interactions that indirectly support neoplastic cell viability and progression. Further evidence also suggests that autocrine TGF β signaling is operative in some tumor cells and can also contribute to tumor invasion, survival, and metastases. These multiple tumor-permissive effects of TGF β provide for a therapeutic opportunity in that by blocking this signaling network, one can interrupt multiple autocrine and paracrine mechanisms that are essential for tumor maintenance. Many questions exist about the viability of this treatment strategy as it applies to patient and tumor selection, molecular predictors of response to therapy, biochemical surrogates for determination of optimal dosing, potential toxicities, and the possible inclusion of inhibitors of TGF β activation to the strategies summarized above. Nonetheless, the plethora of TGF β antagonists in development suggest that these questions will be addressed in the near future.

Acknowledgments

Supported by R01 CA62212 (C.L.A.), Breast Cancer Specialized Program of Research Excellence (SPORE) Grant P50 CA98131, and Vanderbilt-Ingram Cancer Center Support Grant CA68485. We apologize to those colleagues whose work is not referenced because of space considerations or our oversight.

References

- Ananth, S., Knebelmann, B., Gruning, W., Dhanabal, M., Walz, G., Stillman, I.E., and Sukhatme, V.P. (1999). Transforming growth factor $\beta 1$ is a target for the von Hippel-Lindau tumor suppressor and a critical growth factor for clear cell renal carcinoma. *Cancer Res.* 59, 2210–2216.
- Arteaga, C.L., Carty-Dugger, T., Moses, H.L., Hurd, S.D., and Pietenpol, J.A. (1993a). Transforming growth factor $\beta 1$ can induce estrogen-independent tumorigenicity of human breast cancer cells in athymic mice. *Cell Growth Differ.* 4, 193–201.
- Arteaga, C.L., Hurd, S.D., Winnier, A.R., Johnson, M.D., Fendly, B.M., and Forbes, J.T. (1993b). Anti-transforming growth factor (TGF)- β antibodies inhibit breast cancer cell tumorigenicity and increase mouse spleen natural killer cell activity. Implications for a possible role of tumor cell/host TGF- β interactions in human breast cancer progression. *J. Clin. Invest.* 92, 2569–2576.
- Bakin, A.V., Tomlinson, A.K., Bhowmick, N.A., Moses, H.L., and Arteaga, C.L. (2000). Phosphatidylinositol 3-kinase function is required for transforming growth factor β -mediated epithelial to mesenchymal transition and cell migration. *J. Biol. Chem.* 275, 36803–36810.
- Bandyopadhyay, A., Zhu, Y., Cibull, M.L., Bao, L., Chen, C., and Sun, L. (1999). A soluble transforming growth factor β type III receptor suppresses tumorigenicity and metastasis of human breast cancer MDA-MB-231 cells. *Cancer Res.* 59, 5041–5046.
- Bandyopadhyay, A., Lopez-Casillas, F., Malik, S.N., Montiel, J.L., Mendoza, V., Yang, J., and Sun, L.Z. (2002). Antitumor activity of a recombinant soluble betaglycan in human breast cancer xenograft. *Cancer Res.* 62, 4690–4695.
- Bottinger, E.P., Jakubczak, J.L., Haines, D.C., Bagnall, K., and Wakefield, L.M. (1997). Transgenic mice overexpressing a dominant-negative mutant type II transforming growth factor β receptor show enhanced tumorigenesis in the mammary gland and lung in response to the carcinogen 7,12-dimethylbenz-[a]-anthracene. *Cancer Res.* 57, 5564–5570.
- Cui, W., Fowles, D.J., Bryson, S., Duffie, E., Ireland, H., Balmain, A., and Akhurst, R.J. (1996). TGF $\beta 1$ inhibits the formation of benign skin tumors, but enhances progression to invasive spindle carcinomas in transgenic mice. *Cell* 86, 531–542.
- Derynck, R., Akhurst, R.J., and Balmain, A. (2001). TGF- β signaling in tumor suppression and cancer progression. *Nat. Genet.* 29, 117–129.
- Dumont, N., and Arteaga, C.L. (2000). Transforming growth factor- β and breast cancer: Tumor promoting effects of transforming growth factor- β . *Breast Cancer Res.* 2, 125–132.
- Dumont, N., Bakin, A.V., and Arteaga, C.L. (2003). Autocrine transforming growth factor- β signaling mediates Smad-independent motility in human cancer cells. *J. Biol. Chem.* 278, 3275–3285.
- Engle, S.J., Hoying, J.B., Boivin, G.P., Ormsby, I., Gartside, P.S., and Doetschman, T. (1999). Transforming growth factor $\beta 1$ suppresses non-metastatic colon cancer at an early stage of tumorigenesis. *Cancer Res.* 59, 3379–3386.
- Gobbi, H., Dupont, W.D., Simpson, J.F., Plummer, W.D., Jr., Schuyler, P.A., Olson, S.J., Arteaga, C.L., and Page, D.L. (1999). Transforming growth factor- β and breast cancer risk in women with mammary epithelial hyperplasia. *J. Natl. Cancer Inst.* 91, 2096–2101.
- Gohongi, T., Fukumura, D., Boucher, Y., Yun, C.O., Soff, G.A., Compton, C., Todoroki, T., and Jain, R.K. (1999). Tumor-host interactions in the gallbladder suppress distal angiogenesis and tumor growth: involvement of transforming growth factor $\beta 1$. *Nat. Med.* 5, 1203–1208.
- Gorelik, L., and Flavell, R.A. (2001). Immune-mediated eradication of tumors through the blockade of transforming growth factor- β signaling in T cells. *Nat. Med.* 7, 1118–1122.
- Gorska, A.E., Joseph, H., Derynck, R., Moses, H.L., and Serra, R. (1998). Dominant-negative interference of the transforming growth factor β type II receptor in mammary gland epithelium results in alveolar hyperplasia and differentiation in virgin mice. *Cell Growth Differ.* 9, 229–238.
- Hahn, S.A., Schutte, M., Hoque, A.T., Moskaluk, C.A., da Costa, L.T., Rozenblum, E., Weinstein, C.L., Fischer, A., Yeo, C.J., Hruban, R.H., and Kern, S.E. (1996). DPC4, a candidate tumor suppressor gene at human chromosome 18q21.1. *Science* 271, 350–353.
- Iglesias, M., Frontelo, P., Gamallo, C., and Quintanilla, M. (2000). Blockade of Smad4 in transformed keratinocytes containing a Ras oncogene leads to hyperactivation of the Ras-dependent Erk signalling pathway associated with progression to undifferentiated carcinomas. *Oncogene* 19, 4134–4145.
- Janda, E., Lehmann, K., Killisch, I., Jechlinger, M., Herzig, M., Downward, J., Beug, H., and Grunert, S. (2002). Ras and TGF β cooperatively regulate epithelial cell plasticity and metastasis: dissection of Ras signaling pathways. *J. Cell Biol.* 156, 299–313.
- Kleeff, J., Ishiwa, T., Maruyama, H., Friess, H., Truong, P., Buchler, M.W., Falb, D., and Korc, M. (1999). The TGF- β signaling inhibitor Smad7 enhances tumorigenicity in pancreatic cancer. *Oncogene* 18, 5363–5372.
- Kuppner, M.C., Hamou, M.F., Bodmer, S., Fontana, A., and de Tribolet, N. (1988). The glioblastoma-derived T-cell suppressor factor/transforming growth factor $\beta 2$ inhibits the generation of lymphokine-activated killer (LAK) cells. *Int. J. Cancer* 42, 562–567.
- Laping, N.J., Grygielko, E., Mathur, A., Butter, S., Bomberger, J., Tweed, C., Martin, W., Fornwald, J., Lehr, R., Harling, J., et al. (2002). Inhibition of transforming growth factor (TGF)- $\beta 1$ -induced extracellular matrix with a novel inhibitor of the TGF- β type I receptor kinase activity: SB-431542. *Mol. Pharmacol.* 62, 58–64.
- Markowitz, S., Wang, J., Myeroff, L., Parsons, R., Sun, L., Lutterbaugh, J., Fan, R.S., Zborowska, E., Kinzler, K.W., Vogelstein, B., et al. (1995). Inactivation of the type II TGF- β receptor in colon cancer cells with microsatellite instability. *Science* 268, 1336–1338.
- Marzo, A.L., Fitzpatrick, D.R., Robinson, B.W., and Scott, B. (1997). Antisense oligonucleotides specific for transforming growth factor $\beta 2$ inhibit the growth of malignant mesothelioma both in vitro and in vivo. *Cancer Res.* 57, 3200–3207.
- Massagué, J. (1998). TGF- β signal transduction. *Annu. Rev. Biochem.* 67, 753–791.
- Massagué, J., Blain, S.W., and Lo, R.S. (2000). TGF β signaling in growth control, cancer, and heritable disorders. *Cell* 103, 295–309.
- Miettinen, P.J., Ebner, R., Lopez, A.R., and Derynck, R. (1994). TGF- β induced transdifferentiation of mammary epithelial cells to mesenchymal cells: involvement of type I receptors. *J. Cell Biol.* 127, 2021–2036.
- Muraoka, R.S., Dumont, N., Ritter, C.A., Dugger, T.C., Brantley, D.M., Chen, J., Easterly, E., Roebuck, L.R., Ryan, S., Gotwals, P.J., et al. (2002). Blockade of TGF- β inhibits mammary tumor cell viability, migration, and metastases. *J. Clin. Invest.* 109, 1551–1559.
- Nakao, A., Afrakhte, M., Moren, A., Nakayama, T., Christian, J.L., Heuchel, R., Itoh, S., Kawabata, M., Heldin, N.E., Heldin, C.H., and ten Dijke, P. (1997). Identification of Smad7, a TGF β -inducible antagonist of TGF- β signalling. *Nature* 389, 631–635.
- Oft, M., Peli, J., Rudaz, C., Schwarz, H., Beug, H., and Reichmann, E. (1996). TGF- $\beta 1$ and Ha-Ras collaborate in modulating the phenotypic plasticity and invasiveness of epithelial tumor cells. *Genes Dev.* 10, 2462–2477.
- Oft, M., Heider, K.H., and Beug, H. (1998). TGF β signaling is necessary for carcinoma cell invasiveness and metastasis. *Curr. Biol.* 8, 1243–1252.
- Oft, M., Akhurst, R.J., and Balmain, A. (2002). Metastasis is driven by sequential elevation of H-ras and Smad2 levels. *Nat. Cell Biol.* 4, 487–494.
- Pierce, D.F., Jr., Johnson, M.D., Matsui, Y., Robinson, S.D., Gold, L.I., Purchio, A.F., Daniel, C.W., Hogan, B.L., and Moses, H.L. (1993). Inhibition of mammary duct development but not alveolar outgrowth during pregnancy in transgenic mice expressing active TGF- $\beta 1$. *Genes Dev.* 7, 2308–2317.
- Pierce, D.F., Jr., Gorska, A.E., Chytil, A., Meise, K.S., Page, D.L., Coffey, R.J., Jr., and Moses, H.L. (1995). Mammary tumor suppression by transforming growth factor $\beta 1$ transgene expression. *Proc. Natl. Acad. Sci. USA* 92, 4254–4258.
- Rowland-Goldsmith, M.A., Maruyama, H., Kusama, T., Ralli, S., and Korc, M. (2001). Soluble type II transforming growth factor- β (TGF- β) receptor inhibits TGF- β signaling in COLO-357 pancreatic cancer cells in vitro and attenuates tumor formation. *Clin. Cancer Res.* 7, 2931–2940.

- Sawyer, J.S., Anderson, B.D., Beight, D.W., Campbell, R.M., Jones, M.L., Herron, D.K., Lampe, J.W., McCowan, J.R., McMillen, W.T., Mort, N., et al. (2003). Synthesis and activity of new aryl- and heteroaryl-substituted pyrazole inhibitors of the transforming growth factor- β type I receptor kinase domain. *J. Med. Chem.*, in press.
- Shin, I., Bakin, A.V., Rodeck, U., Brunet, A., and Arteaga, C.L. (2001). Transforming growth factor beta enhances epithelial cell survival via Akt-dependent regulation of FKHRL1. *Mol. Biol. Cell* 12, 3328–3339.
- Siriwardena, D., Khaw, P.T., King, A.J., Donaldson, M.L., Overton, B.M., Migdal, C., and Cordeiro, M.F. (2002). Human antitransforming growth factor β (2) monoclonal antibody—a new modulator of wound healing in trabeculectomy: a randomized placebo controlled clinical study. *Ophthalmology* 109, 427–431.
- Tang, B., Bottinger, E.P., Jakowlew, S.B., Bagnall, K.M., Mariano, J., Anver, M.R., Letterio, J.J., and Wakefield, L.M. (1998). Transforming growth factor- β 1 is a new form of tumor suppressor with true haploid insufficiency. *Nat. Med.* 4, 802–807.
- Tsushima, H., Ito, N., Tamura, S., Matsuda, Y., Inada, M., Yabuuchi, I., Imai, Y., Nagashima, R., Misawa, H., Takeda, H., et al. (2001). Circulating transforming growth factor β 1 as a predictor of liver metastasis after resection in colorectal cancer. *Clin. Cancer Res.* 7, 1258–1262.
- Wagner, M., Kleeff, J., Friess, H., Buchler, M.W., and Korc, M. (1999). Enhanced expression of the type II transforming growth factor- β receptor is associated with decreased survival in human pancreatic cancer. *Pancreas* 19, 370–376.
- Watanabe, T., Wu, T.T., Catalano, P.J., Ueki, T., Satriano, R., Haller, D.G., Benson, A.B., 3rd, and Hamilton, S.R. (2001). Molecular predictors of survival after adjuvant chemotherapy for colon cancer. *N. Engl. J. Med.* 344, 1196–1206.
- Weeks, B.H., He, W., Olson, K.L., and Wang, X.J. (2001). Inducible expression of transforming growth factor beta1 in papillomas causes rapid metastasis. *Cancer Res.* 61, 7435–7443.
- Wojtowicz-Praga, S. (2003). Reversal of tumor-induced immunosuppression with TGF- β inhibitors. *Invest. New Drugs* 21, 1–12.
- Wu, S.P., Sun, L.Z., Willson, J.K., Humphrey, L., Kerbel, R., and Brattain, M.G. (1993). Repression of autocrine transforming growth factor β 1 and β 2 in quiescent CBS colon carcinoma cells leads to progression of tumorigenic properties. *Cell Growth Differ.* 4, 115–123.
- Xu, X., Brodie, S.G., Yang, X., Im, Y.H., Parks, W.T., Chen, L., Zhou, Y.X., Weinstein, M., Kim, S.J., and Deng, C.X. (2000). Haploid loss of the tumor suppressor Smad4/Dpc4 initiates gastric polyposis and cancer in mice. *Oncogene* 19, 1868–1874.
- Yang, Y.A., Dukhanina, O., Tang, B., Mamura, M., Letterio, J.J., MacGregor, J., Patel, S.C., Khozin, S., Liu, Z.Y., Green, J., et al. (2002). Lifetime exposure to a soluble TGF- β antagonist protects mice against metastasis without adverse side effects. *J. Clin. Invest.* 109, 1607–1615.
- Zhu, Y., Richardson, J.A., Parada, L.F., and Graff, J.M. (1998). Smad3 mutant mice develop metastatic colorectal cancer. *Cell* 94, 703–714.
- Ziv, E., Cauley, J., Morin, P.A., Saiz, R., and Browner, W.S. (2001). Association between the T29→C polymorphism in the transforming growth factor β 1 gene and breast cancer among elderly white women: The study of osteoporotic fractures. *JAMA* 285, 2859–2863.
- Zugmaier, G., Paik, S., Wilding, G., Knabbe, C., Bano, M., Lupu, R., Deschauer, B., Simpson, S., Dickson, R.B., and Lippman, M. (1991). Transforming growth factor β 1 induces cachexia and systemic fibrosis without an antitumor effect in nude mice. *Cancer Res.* 51, 3590–3594.